

Novocastra™ Liquid Mouse Monoclonal Antibody Mismatch Repair Protein (MLH1)

Product Code: NCL-L-MLH1

Intended Use	FOR RESEARCH USE ONLY.
Specificity	Human MLH1 antigen.
Clone	ES05
Ig Class	IgG1 (Kappa)
Antigen Used for Immunizations	Prokaryotic recombinant protein corresponding to a 210 amino acid portion of the human MLH1 molecule.
Hybridoma Partner	Mouse myeloma (p3-NS1-Ag4-1).
Preparation	A purified immunoglobulin fraction diluted in PBS with 1% BSA containing 15 mM sodium azide as a preservative. Volume as indicated on vial label.
Effective on Frozen Tissue	Not evaluated.
Effective on Paraffin Wax Embedded Tissue	Yes (using high temperature antigen unmasking technique: see overleaf).
Recommendations on Use	Immunohistochemistry: Typical working dilution 1:50. Citrate based buffer, pH 6.0. 30 minutes primary antibody incubation at 25 °C. Polymer detection recommended. Western Blotting: Typical working dilution 1: 2000–4000 (ECL™, Amersham Pharmacia Biotech).
Positive Controls	Immunohistochemistry: Colon. Western Blotting: Jurkat cells.
Staining Pattern	Nuclear.
Storage and Stability	Store liquid antibody at 4 °C. Under these conditions, there is no significant loss in product performance up to the expiry date indicated on the vial label. Prepare working dilutions on the day of use.

General Overview	MLH1 is a mismatch repair protein involved in maintaining the integrity of genetic information, alongside MSH2, MSH6 and PMS2. During DNA replication, strand misalignment can occur resulting in alterations to microsatellite repeats, often referred to as microsatellite instability (MSI). These defects in DNA repair pathways have been linked to human carcinogenesis. Mutations in the MLH1 gene have been reported to be found in tumors with MSI, such as some forms of colon cancer e.g. Hereditary nonpolyposis colon cancer (HNPCC), a subset of sporadic carcinomas and breast cancer. Loss of expression of MLH1 has also been reported in acute lymphoblastic leukemias, endometrial carcinomas, gastric carcinomas and ovarian carcinomas.
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General References	Matheson EC, Hall AG. <i>Carcinogenesis</i> . 24(1): 31–38 (2003). Castrilli G, Fabiano A, La Torre G et al. <i>Journal of Oral Pathology and Medicine</i> . 31(4): 234–238 (2002). Ichikawa Y, Tsunoda H, Takano K et al. <i>Journal of Clinical Oncology</i> . 32: 110–112 (2002). Machin P, Catusas L, Pons C et al. <i>Journal of Cutaneous Pathology</i> . 29: 415–420 (2002). Murata H, Khattar NH, Kang Y et al. <i>Oncogene</i> . 21(37): 5696–5703 (2002). Ruszkiewicz A, Bennett G, Moore J et al. <i>Pathology</i> . 34(6): 541–547 (2002). Teruya-Feldstein J, Greene J, Cohen L et al. <i>Leukemia and Lymphoma</i> . 43(8): 1619–1626 (2002). Murata H, Khattar NH, Kang Y et al. <i>Oncogene</i> . 21(37): 5696–5703 (2002). Perrin J, Gouvernet J, Parriaux D et al. <i>Journal of Oncology</i> . 19: 891–895 (2001). Bronner CE, Baker SM, Morrison PT et al. <i>Nature</i> . 368: 258–261(1994).
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Instructions for Use

High Temperature Antigen Unmasking Technique for Immunohistochemical Demonstration on Paraffin Sections

1. Cut and mount sections on slides coated with a suitable tissue adhesive.
2. Deparaffinize sections and rehydrate to distilled water.
3. Place sections in 0.5% hydrogen peroxide/methanol for 10 minutes (or use other appropriate endogenous peroxidase blocking procedure). Wash sections in tap water.
4. Heat 1500 mL of the recommended unmasking solution (0.01 M citrate buffer, pH 6.0 (or Epitope Retrieval Solution, RE7113) unless otherwise indicated overleaf) until boiling in a stainless steel pressure cooker. Cover but do not lock lid.
5. Position slides into metal staining racks (do not place slides close together as uneven staining may occur) and lower into pressure cooker ensuring slides are completely immersed in unmasking solution. Lock lid.
6. When the pressure cooker reaches operating temperature and pressure (after about 5 minutes) start a timer for 1 minute (unless otherwise indicated on the data sheet).
7. When the timer rings, remove pressure cooker from heat source and run under cold water with lid on. DO NOT OPEN LID UNTIL THE INDICATORS SHOW THAT PRESSURE HAS BEEN RELEASED. Open lid, remove slides and place immediately into a bath of tap water.
8. Wash sections in TBS* buffer (pH 7.6) for 1 x 5 minutes.
9. Place sections in diluted normal serum (or RTU Normal Horse Serum) for 10 minutes.
10. Incubate sections with primary antibody. Use Antibody Diluent RE7133 (where available).
11. Wash in TBS buffer for 2 x 5 minutes.
12. Incubate sections in an appropriate biotinylated secondary antibody.
13. Wash in TBS buffer for 2 x 5 minutes.
14. Incubate slides in ABC reagent (or RTU streptavidin/peroxidase complex).
15. Wash in TBS buffer for 2 x 5 minutes.
16. Incubate slides in DAB or other suitable peroxidase substrate.
17. Wash thoroughly in running tap water.
18. Counterstain with hematoxylin (if required), dehydrate and mount.

Solutions

0.01 M CITRATE BUFFER (pH 6.0) or RE7113 (where available).

Add 3.84 g of citric acid (anhydrous) to 1.8 L of distilled water. Adjust to pH 6.0 using concentrated NaOH. Make up to 2 L with distilled water.

1 mM EDTA (pH 8.0) or RE7116 (where available).

Add 0.37 g of EDTA (SIGMA product code E-5134) to 1 litre of distilled water. Adjust pH to 8.0 using 1.0 M NaOH.

20 mM TRIS/ 0.65 mM EDTA/ 0.005% TWEEN (pH 9.0) or RE7119 (where available).

Dissolve 14.4 g Tris (BDH product code 271197K) and 1.44 g EDTA (SIGMA product code E-5134) to 0.55 L of distilled water. Adjust pH to 9.0 with 1 M HCl and add 0.3 mL Tween 20 (SIGMA product code P-1379). Make up to 0.6 L with distilled water. This is a 10x concentrate which should be diluted with distilled water as required (eg 150 mL diluted with 1350 mL of distilled water).

* In most applications, 10 mM phosphate, 0.15 M NaCl, pH 7.6 (PBS) can be used instead of 50 mM Tris, 0.15 M NaCl, pH 7.6 (TBS).

Safety Note

To ensure the correct and safe use of your pressure cooker, PLEASE READ MANUFACTURER'S INSTRUCTIONS.