

# Novocastra™ Liquid Mouse Monoclonal Antibody Alpha-Sarcoglycan (Adhalin)

**Product Code: NCL-L-a-SARC**

<b>Intended Use</b>	FOR RESEARCH USE ONLY.
<b>Specificity</b>	Human alpha-sarcoglycan, also known as adhalin. Also crossreacts strongly with alpha-sarcoglycan in sections of muscle from mouse, rat, rabbit, hamster and pig. Does not react with chicken muscle.
<b>Clone</b>	Ad1/20A6
<b>Ig Class</b>	IgG1
<b>Antigen Used for Immunizations</b>	Fusion protein containing amino acids 217 to 289 of the rabbit adhalin sequence (The Journal of Biological Chemistry. 268: 23739–23742, 1993).
<b>Hybridoma Partner</b>	Mouse myeloma (X63.Ag8.653) x BKT0.
<b>Preparation</b>	Liquid tissue culture supernatant containing 15 mM sodium azide. Volume as indicated on vial label.
<b>Effective on Frozen Tissue</b>	Yes - unfixed.
<b>Effective on Paraffin Wax Embedded Tissue</b>	No
<b>Recommendations on Use</b>	Immunohistochemistry: Typical working dilution 1:100–1:200. 60 minutes primary antibody incubation at 25 °C. Indirect immunoperoxidase technique (see overleaf). Western Blotting: Typical working dilution 1:50–1:100. Standard homogenates, no need to isolate membranes). Electron microscopy gold: Light fixation with 2% formaldehyde + 0.001% glutaraldehyde for 1 hour, 2.3 M sucrose used as a cryoprotectant is recommended. Typical working dilution NEAT. 90 minutes primary antibody incubation at 25 °C.
<b>Positive Controls</b>	Immunohistochemistry: Normal human striated muscle frozen in isopentane chilled in liquid nitrogen. Western Blotting: Skeletal muscle. Electron microscopy gold: Normal human striated muscle.
<b>Staining Pattern</b>	Light microscope: continuous labelling around the muscle fiber membranes. Western blotting: band at approximately 50 kD, in muscle extracts. Electron microscopy gold: at the plasma membrane of muscle fibers.
<b>Storage and Stability</b>	Store liquid antibody at 4 °C. Under these conditions, there is no significant loss in product performance up to the expiry date indicated on the vial label. Prepare working dilutions on the day of use.
<b>General Overview</b>	In normal skeletal muscle, dystrophin is attached to the muscle membrane via a complex of at least seven proteins (dystrophin associated glycoproteins, DAGs). The biological significance of this dystrophin/glycoprotein complex is not fully understood, but it appears to form an essential linkage between actin on the inside of the muscle fiber and muscle laminin in the basal lamina which surrounds the fiber. Labeling with an antibody to beta-spectrin, eg NCL-SPEC1 (recommended for human use), to monitor membrane integrity, is an essential immunohistochemical control in any research performed.
<b>General References</b>	Sheriffs I N, Rampling D and Smith V V. Journal of Clinical Pathology. 54: 517–520 (2001). Picolo F, Roberds S L, Jeanpierre M, et al.. Nature Genetics. 10: 243–245 (1995). Roberds S L, Anderson R D, Ibraghimov-Beskrovnaia O, et al.. The Journal of Biological Chemistry. 268: 23739–23742 (1993). Matsumura K, Tomé F M S, Collin H, et al.. Nature. 359: 320–322 (1992). Ervasti J M and Campbell K P. Cell. 66: 1121–1131 (1991).



## Instructions for Use

**Protocol for Immunohistochemical use  
of the following Monoclonal Antibodies:  
NCL-alpha-ACT, NCL-a-SARC, NCL-b-  
SARC, NCL-d-SARC, NCL-g-SARC, NCL-  
b-DG,  
NCL-MHCd, NCL-MHCf, NCL-MHCn,  
NCL-MHCs, NCL-SPEC1, NCL-SPEC2,  
NCL-DRP2, NCL-MEROSIN,  
NCL-Hamlet and NCL-Hamlet-2.**

1. Freeze muscle blocks in isopentane chilled in liquid nitrogen.
2. Cut 4–10 µm sections and air dry on slides coated with tissue adhesive.
3. Slides may be stored below -70 °C wrapped in cling film until required. If stored sections are used, allow sections to equilibrate to 25 °C before unwrapping and proceeding.
4. Apply a 50 µl aliquot of primary antibody to section (unfixed) Use Antibody Diluent RE7133 (where available). Incubate for 1 hour at 25 °C or 37 °C.  
**Please note** that where NCL-Hamlet and NCL-Hamlet-2 primary antibodies are used, it is recommended that sections are fixed in acetone/methanol (1:1) for 4 minutes at room temperature prior to incubation with the primary antibody.
5. Wash sections in TBS\* buffer (pH 7.6) for 3 x 10 minutes.
6. Apply a 50 µL aliquot of labeled secondary antibody (e.g. NCL-GAMP diluted 1:100). Incubate for 1 hour at 25 °C.
7. Wash sections in TBS\* buffer (pH 7.6) for 3 x 10 minutes.
8. Mount fluorescent sections in aqueous mountant or visualize peroxidase label (e.g. by exposure to freshly prepared 0.05% w/v diaminobenzidine in TBS\* buffer containing 0.1% w/v hydrogen peroxide). Dehydrate, clear and mount peroxidase labeled sections for permanent preparations.

\* In most applications, 10 mM phosphate, 0.15 M NaCl, pH 7.6 (PBS) can be used instead of 50 mM Tris, 0.15 M NaCl, pH 7.6 (TBS).