Novocastra™ Lyophilized Mouse Monoclonal Antibody Delta-Sarcoglycan

Product Code: NCL-d-SARC

Intended Use	FOR RESEARCH USE ONLY.
Specificity	Human delta-sarcoglycan (35 kD). Does not react with delta-sarcoglycan in sections of mouse, rat, rabbit, dog, chicken, hamster or pig muscle. Other animal species not tested.
Clone	δSarc3/12C1
Ig Class	IgG2a, kappa
Antigen Used for Immunizations	Synthetic peptide containing amino acids 1–19 at the N-terminus of the human δ-sarcoglycan sequence (Jung D, et al., 1996).
Hybridoma Partner	Mouse myeloma (X63.Ag8.653) x CD1.
Preparation	Lyophilized tissue culture supernatant containing 15 mM sodium azide. Reconstitute with the volume of sterile distilled water indicated on the vial label.
Effective on Frozen Tissue	Yes - unfixed.
Effective on Paraffin Wax Embedded Tissue	Not evaluated.
Recommendations on Use	Immunohistochemistry: Typical working dilution 1:25–1:50. 60 minutes primary antibody incubation at 25 °C. Indirect immunoperoxidase technique (see overleaf). Western Blotting: Typical working dilution 1:25–1:50.
Positive Controls	Immunohistochemistry: Normal human striated muscle frozen in isopentane chilled in liquid nitrogen. Western Blotting: Muscle
Staining Pattern	Light microscope: continuous labelling around normal muscle fibre membranes.
Storage and Stability	Store unopened lyophilized antibody at 4 °C. Under these conditions, there is no significant loss in product performance up to the expiry date indicated on the vial label. The reconstituted antibody is stable for at least two months when stored at 4 °C. For long term storage, it is recommended that aliquots of the antibody are frozen at -20 °C (frost-free freezers are not recommended). Repeated freezing and thawing must be avoided. Prepare working dilutions on the day of use.
General Overview	In normal skeletal muscle, dystrophin, the protein product of the gene which is defective in Duchenne and Becker muscular dystrophy, is attached to the muscle membrane via a complex of at least seven proteins (dystrophin associated glycoproteins, DAGs) including the 4 sarcoglycans (alpha, beta, gamma and delta). Dystrophin-deficient muscle shows a generalised reduction in DAG labelling. Important: For reliable interpretation of dystrophin labelling patterns using tissue sections, the use of a SPECTRIN control is essential.
General References	Sheriffs I N, Rampling D and Smith V V. Journal of Clinical Pathology. 54: 517–520 (2001). Jung D, Duclos F, Aposol B, et al.. The Journal of Biological Chemistry. 271 (50): 32321–32329 (1996).

Leica Biosystems Newcastle Ltd
Balliol Business Park West
Benton Lane
Newcastle Upon Tyne NE12 8EW
United Kingdom
✆ +44 191 215 4242

Instructions for Use

Protocol for Immunohistochemical use of the following Monoclonal Antibodies: NCL-alpha-ACT, NCL-a-SARC, NCL-b-SARC, NCL-d-SARC, NCL-g-SARC, NCL-b-DG, NCL-MHCd, NCL-MHCf, NCL-MHCn, NCL-MHCs, NCL-SPEC1, NCL-SPEC2, NCL-DRP2, NCL-MEROSIN, NCL-Hamlet and NCL-Hamlet-2.

1. Freeze muscle blocks in isopentane chilled in liquid nitrogen.
2. Cut 4–10 µm sections and air dry on slides coated with tissue adhesive.
3. Slides may be stored below -70 °C wrapped in cling film until required. If stored sections are used, allow sections to equilibriate to 25 °C before unwrapping and proceeding.
4. Apply a 50 µl aliquot of primary antibody to section (unfixed) Use Antibody Diluent RE7133 (where available). Incubate for 1 hour at 25 °C or 37 °C.
   Please note that where NCL-Hamlet and NCL-Hamlet-2 primary antibodies are used, it is recommended that sections are fixed in acetone/methanol (1:1) for 4 minutes at room temperature prior to incubation with the primary antibody.
5. Wash sections in TBS* buffer (pH 7.6) for 3 x 10 minutes.
6. Apply a 50 µL aliquot of labeled secondary antibody (e.g. NCL-GAMP diluted 1:100). Incubate for 1 hour at 25 °C.
7. Wash sections in TBS* buffer (pH 7.6) for 3 x 10 minutes.
8. Mount fluorescent sections in aqueous mountant or visualize peroxidase label (e.g. by exposure to freshly prepared 0.05% w/v diaminobenzidine in TBS* buffer containing 0.1% w/v hydrogen peroxide). Dehydrate, clear and mount peroxidase labeled sections for permanent preparations.

** In most applications, 10 mM phosphate, 0.15 M NaCl, pH 7.6 (PBS) can be used instead of 50 mM Tris, 0.15 M NaCl, pH 7.6 (TBS).*