

Novocastra™ Lyophilized Mouse Monoclonal Antibody Dystrophin (Rod Domain)



Product Code: NCL-DYS1

Intended Use	FOR RESEARCH USE ONLY.
Specificity	Reacts strongly with the mid rod domain (between amino acids 1181 and 1388) of human dystrophin. Also reacts with skeletal, cardiac and smooth muscle dystrophin from normal mouse, rat, rabbit, dog, hamster and pig. Reacts on blots with the brain isoform. No reactivity with mdx mouse tissue of DMD/BMD patients who have a gene deletion which removes the antibody binding site. No reaction is observed with chicken dystrophin.
Clone	Dy4/6D3
Ig Class	IgG2a
Antigen Used for Immunizations	Bacterial fusion protein. (Hoffman et al., 1987).
Hybridoma Partner	Mouse myeloma (X63.Ag8.653) x CD1.
Preparation	Lyophilized tissue culture supernatant containing 15 mM sodium azide. Reconstitute with the volume of sterile distilled water indicated on the vial label.
Effective on Frozen Tissue	Yes - unfixed
Effective on Paraffin Wax Embedded Tissue	No
Recommendations on Use	Immunohistochemistry: Typical working dilution 1:20. Indirect immunoperoxidase technique (see overleaf). Western Blotting: Typical working dilution 1:100–1:250. Electron microscopy gold: Light fixation with 2% formaldehyde + 0.001% glutaraldehyde for 1 hour, 2.3 M sucrose used as cryoprotectant is recommended. Typical working dilution NEAT. 90 minutes primary antibody incubation at 25 °C.
Positive Controls	Immunohistochemistry - Snap frozen normal human or rat striated muscle. Western Blotting - Skeletal muscle. Electron microscopy gold - Snap frozen normal human or rat striated muscle.
Staining Pattern	Light microscope: continuous rim of labelling at the periphery of muscle fibres. Western blotting: strong doublet of bands at approximately 400 kD plus metabolites of lower molecular mass. Electron microscopy gold: close to the cytoplasmic face of the plasma membrane.
Storage and Stability	Store unopened lyophilized antibody at 4 °C. Under these conditions, there is no significant loss in product performance up to the expiry date indicated on the vial label. The reconstituted antibody is stable for at least two months when stored at 4 °C. For long term storage, it is recommended that aliquots of the antibody are frozen at -20 °C (frost-free freezers are not recommended). Repeated freezing and thawing must be avoided. Prepare working dilutions on the day of use.
General Overview	Duchenne muscular dystrophy (DMD) is the most severe of the muscular dystrophies resulting in progressive muscular wasting and death. Dystrophin is the 427 kD protein product of the Duchenne Muscular Dystrophy/Becker Muscular Dystrophy (DMD/BMD) gene located on the X chromosome at position Xp21. Western blotting and immunohistochemistry are the two established methods for use in research studies for the detection of abnormalities of dystrophin expression in muscle biopsies. Important: For reliable interpretation of dystrophin labelling patterns using tissue sections, the use of a SPECTRIN control is essential.
General References	Sheriffs I N, Rampling D and Smith V V. Journal of Clinical Pathology, 54: 517–520 (2001). Hoffman E P, Brown R H Jr and Kunkell L M. Cell. 51 (6): 919–928 (1987).



Instructions for Use

Protocol for Immunohistochemical Use of Monoclonal Antibodies to Dystrophin; NCL-DYS1, NCL-DYS2 and NCL-DYS3

1. Freeze muscle blocks in isopentane chilled in liquid nitrogen.
2. Cut 4–10 µm sections and air dry on slides coated with tissue adhesive.
3. Slides may be stored at -70 °C wrapped in cling film until required. If stored sections are used, allow sections to equilibrate to room temperature before unwrapping and proceeding.
4. Apply a 50 µL aliquot of primary antibody to section (unfixed). Incubate for 1 hour at 25 °C or 37 °C.
5. Wash sections 3 x 10 minutes in phosphate buffered saline.
6. Apply a 50 µL aliquot of labelled secondary antibody. Incubate for 1 hour at 25 °C.
7. Wash sections 3 x 10 minutes in phosphate buffered saline.
8. Mount fluorescent sections in aqueous mountant or visualize peroxidase label (e.g. by exposure to freshly prepared 0.05% w/v diaminobenzidine in phosphate buffered saline containing 0.1% w/v hydrogen peroxide). Dehydrate, clear and mount peroxidase labelled sections for permanent preparations.

References

- Nicholson L V B, Johnson M A, Danson K, et al.. Dystrophin or a 'related protein' in Duchenne muscular dystrophy. *Acta Neurol. Scand.* 86: 8–14 (1992).
- Slater C R and Nicholson L V B. Is dystrophin labelling always discontinuous in Becker muscular dystrophy? *Journal of Neurological Sciences.* 101: 187–192 (1991).
- Voit T, Stuetgen P, Cremer M, et al.. Dystrophin as a diagnostic marker in Duchenne and Becker muscular dystrophy. Correlation of immunofluorescence and western blot. *Neuropediatrics.* 22: 152–162 (1991).
- Cullen M H, Walsh J, Nicholson L V B, et al.. Ultrastructural localisation of dystrophin in human muscle by using gold immunolabelling. *Proceedings of the Royal Society.* 240(B): 197–210 (1990).
- England S B, Nicholson L V B, Johnson M A, et al.. Very mild muscular dystrophy associated with the deletion of 46% of dystrophin. *Nature.* 324: 180–182 (1990).
- Nicholson L V B, Johnson M A, Gardner-Medwin D, et al.. Heterogeneity of dystrophin expression in patients with Duchenne and Becker muscular dystrophy. (226 patient study. Paper contains details about the combined use of immunocytochemistry and Western blotting, and provides guidelines for differential diagnosis using dystrophin protein analysis). *Acta Neuropathologica.* 80: 239–250 (1990).
- Nordenskjold M, Nicholson L V B, Edstrom L, et al.. A normal male with an inherited deletion of one exon within the DMD gene. *Human Genetics.* 84: 207–209 (1990).
- Nicholson L V B, Davison K, Falkous G, et al.. Dystrophin in skeletal muscle. I. Western blot analysis using a monoclonal antibody. *Journal of the Neurological Sciences.* 94: 125–136 (1989).
- Nicholson L V B, Davison K, Johnson M A, et al.. Dystrophin in skeletal muscle. II. Immunoreactivity in patients with Xp21 muscular dystrophy. *Journal of the Neurological Sciences.* 94: 137–146 (1989).
- Norman A M, Hughes H E, Gardner-Medwin D, et al.. Dystrophin analysis in the diagnosis of muscular dystrophy. *Archives of Disease in Childhood.* 64: 1501–1503 (1989).