

Novocastra™ Liquid Mouse Monoclonal Antibody Epidermal Growth Factor Receptor

Product Code: NCL-L-EGFR

Intended Use	FOR RESEARCH USE ONLY.
Specificity	External domain of human epidermal growth factor receptor.
Clone	EGFR.113
Ig Class	IgG2a
Antigen Used for Immunizations	Recombinant fusion protein to the external domain.
Hybridoma Partner	Mouse myeloma (p3-NS1-Ag4-1).
Preparation	Liquid tissue culture supernatant containing 15 mM sodium azide. Volume as indicated on vial label.
Effective on Frozen Tissue	Yes
Effective on Paraffin Wax Embedded Tissue	Yes (using the high temperature antigen unmasking technique: see overleaf).
Recommendations on Use	Immunohistochemistry: Typical working dilution 1:10–1:20. High temperature antigen unmasking technique. 60 minutes primary antibody incubation at 25 °C. Standard ABC technique. Western Blotting: Not recommended.
Positive Controls	Immunohistochemistry: Placenta or normal skin; keratinocytes.
Staining Pattern	Predominant staining is membrane, though cytoplasmic staining may also be observed. Furthermore, in a small proportion of cases, nuclear staining may be seen. This is most likely due to a crossreaction with a nuclear antigen and may be fixation related.
Storage and Stability	Store liquid antibody at 4 °C. Under these conditions, there is no significant loss in product performance up to the expiry date indicated on the vial label. Prepare working dilutions on the day of use.
General Overview	Epidermal growth factor receptor (EGFR) is a 170 kD tyrosine kinase linked protein. EGFR is a member of the type I growth factor family. Activation of the receptor leads to an increase in cellular Ca ²⁺ ions resulting in cellular proliferation. In normal tissues immunoreactivity is limited mainly, to the basal layers of complex epithelia.
General References	Sriplakich S, Jahnon S and Karlsson M G. <i>BJU International</i> . 83 (4): 498–503 (1999). Inoue K, Chikazawa M, Karashima T, et al.. <i>Acta. Med. Okayama</i> . 52 (6): 305–310 (1998). Tungekar M F and Linehan J. <i>Journal of Clinical Pathology</i> . 51: 583–587 (1998). Fox S B, Smith K, Hollyer J, et al.. <i>Breast Cancer Research and Treatment</i> . 29: 41–49 (1994). Harris A L. <i>Breast Cancer Research and Treatment</i> . 29: 1–2 (1994). Rajkumar T and Gullick W J. <i>Breast Cancer Research and Treatment</i> . 29: 3–9 (1994). Nicholson S, Halcrow P, Sainsbury J R C, et al.. <i>British Journal of Cancer</i> . 58: 810–814 (1988). Cerny T, Barnes D M, Hasleton P, et al.. <i>British Journal of Cancer</i> . 54: 265–269 (1986). Damjanov I, Mildner B and Knowles B B. <i>Laboratory Investigations</i> . 55: 588–592 (1986). Gullick W J, Marsden J J, Whittle N, et al.. <i>Cancer Research</i> . 46: 285–292 (1986).



Instructions for Use

High Temperature Antigen Unmasking Technique for Immunohistochemical Demonstration on Paraffin Sections

1. Cut and mount sections on slides coated with a suitable tissue adhesive.
2. Deparaffinize sections and rehydrate to distilled water.
3. Place sections in 0.5% hydrogen peroxide/methanol for 10 minutes (or use other appropriate endogenous peroxidase blocking procedure). Wash sections in tap water.
4. Heat 1500 mL of the recommended unmasking solution (0.01 M citrate buffer, pH 6.0 (or Epitope Retrieval Solution, RE7113) unless otherwise indicated overleaf) until boiling in a stainless steel pressure cooker. Cover but do not lock lid.
5. Position slides into metal staining racks (do not place slides close together as uneven staining may occur) and lower into pressure cooker ensuring slides are completely immersed in unmasking solution. Lock lid.
6. When the pressure cooker reaches operating temperature and pressure (after about 5 minutes) start a timer for 1 minute (unless otherwise indicated on the data sheet).
7. When the timer rings, remove pressure cooker from heat source and run under cold water with lid on. DO NOT OPEN LID UNTIL THE INDICATORS SHOW THAT PRESSURE HAS BEEN RELEASED. Open lid, remove slides and place immediately into a bath of tap water.
8. Wash sections in TBS* buffer (pH 7.6) for 1 x 5 minutes.
9. Place sections in diluted normal serum (or RTU Normal Horse Serum) for 10 minutes.
10. Incubate sections with primary antibody. Use Antibody Diluent RE7133 (where available).
11. Wash in TBS buffer for 2 x 5 minutes.
12. Incubate sections in an appropriate biotinylated secondary antibody.
13. Wash in TBS buffer for 2 x 5 minutes.
14. Incubate slides in ABC reagent (or RTU streptavidin/peroxidase complex).
15. Wash in TBS buffer for 2 x 5 minutes.
16. Incubate slides in DAB or other suitable peroxidase substrate.
17. Wash thoroughly in running tap water.
18. Counterstain with hematoxylin (if required), dehydrate and mount.

Solutions

0.01 M CITRATE BUFFER (pH 6.0) or RE7113 (where available).

Add 3.84 g of citric acid (anhydrous) to 1.8 L of distilled water. Adjust to pH 6.0 using concentrated NaOH. Make up to 2 L with distilled water.

1 mM EDTA (pH 8.0) or RE7116 (where available).

Add 0.37 g of EDTA (SIGMA product code E-5134) to 1 litre of distilled water. Adjust pH to 8.0 using 1.0 M NaOH.

20 mM TRIS/ 0.65 mM EDTA/ 0.005% TWEEN (pH 9.0) or RE7119 (where available).

Dissolve 14.4 g Tris (BDH product code 271197K) and 1.44 g EDTA (SIGMA product code E-5134) to 0.55 L of distilled water. Adjust pH to 9.0 with 1 M HCl and add 0.3 mL Tween 20 (SIGMA product code P-1379). Make up to 0.6 L with distilled water. This is a 10x concentrate which should be diluted with distilled water as required (eg 150 mL diluted with 1350 mL of distilled water).

* In most applications, 10 mM phosphate, 0.15 M NaCl, pH 7.6 (PBS) can be used instead of 50 mM Tris, 0.15 M NaCl, pH 7.6 (TBS).

Safety Note

To ensure the correct and safe use of your pressure cooker, PLEASE READ MANUFACTURER'S INSTRUCTIONS.